

Discovery of Non-Template-Dependent DNA Polymerase

Paper: *Calf Thymus Polymerase*

Author: F.J. Bollum (1960)

Published in: *The Journal of Biological Chemistry*

1. Background and Objective

- The **E. coli DNA polymerase (DNA Polymerase I)** discovered earlier by Kornberg was **template-dependent**, requiring a DNA strand to guide replication.
- The study aimed to **identify and characterize a mammalian DNA polymerase** from calf thymus that could synthesize DNA.
- Unlike Kornberg's enzyme, this polymerase showed **non-template-dependent activity**, meaning it could **synthesize DNA without requiring a complementary strand**.

2. Key Findings

2.1 Purification and Characterization of the Enzyme

- The enzyme, later termed **Terminal Deoxynucleotidyl Transferase (TdT)**, was extracted from **calf thymus** and purified.
- It catalyzed the **addition of deoxyribonucleotides (dNTPs) to the 3'-OH end of a DNA strand**, without using a complementary DNA template.
- The enzyme was distinct from **template-dependent polymerases**, as it lacked the ability to proofread or correct errors.

2.2 Reaction Mechanism

- The polymerase catalyzed the polymerization of **deoxyribonucleoside triphosphates (dNTPs)** in the presence of **magnesium ions (Mg^{2+})**.
- Unlike Kornberg's DNA polymerase I, it **did not require a DNA template** for nucleotide addition.
- The reaction followed a **3' → 5' extension**, forming a **single-stranded DNA tail** at the 3' end of existing DNA molecules.

2.3 Substrate Specificity

- The enzyme required **all four deoxyribonucleotides (dATP, dGTP, dCTP, dTTP)** for maximal activity.
- It showed **variable incorporation rates**, with a preference for **dGTP and dATP** over dCTP and dTTP.
- The polymerase exhibited **higher activity with denatured DNA** rather than native, double-stranded DNA.

2.4 Reverse Reaction: Pyrophosphorolysis

- The enzyme also catalyzed the **reverse reaction** in the presence of **pyrophosphate (PPi)**, leading to the **removal of nucleotides** from the growing DNA chain.
- This demonstrated the **dynamic nature** of the enzyme in DNA metabolism.

3. Experimental Approach

3.1 Extraction and Purification of the Enzyme

The enzyme was isolated from **calf thymus glands** using a series of purification steps:

1. Tissue Homogenization

- Calf thymus glands were **homogenized in a sucrose buffer solution** to extract soluble proteins.
- The extract was **centrifuged** to remove debris, and the supernatant (Fraction A) was collected.

2. Acid Fractionation

- The extract was subjected to **stepwise pH reduction using acetic acid** to precipitate unwanted proteins.
- At **pH 5.9 and pH 5.4**, two distinct precipitates were formed, with **Fraction B** containing the polymerase activity.

3. Ammonium Sulfate Precipitation

- **Selective precipitation** using ammonium sulfate helped concentrate the enzyme further.
- Fraction C contained the **active polymerase** with reduced contaminants.

4. DEAE-Cellulose Chromatography

- Fraction C was passed through a **DEAE-cellulose column**, which removed nucleic acid contaminants and improved purity.
- The enzyme eluted from the column (Fraction D) had **high polymerase activity and minimal DNase contamination**.

3.2 Enzyme Assay: Measuring DNA Synthesis

1. Reaction Setup

- The polymerase was incubated with:
 - **Deoxyribonucleoside triphosphates (dNTPs)**
 - **Magnesium ions (Mg^{2+})**
 - **DNA primers (heat-denatured DNA from calf thymus or salmon sperm DNA)**
- The incorporation of **radioactively labeled thymidine triphosphate (3H -TTP)** into acid-insoluble DNA was measured.

2. Control Experiments

- When **DNA primers were omitted**, no DNA synthesis was detected, confirming that **the enzyme needed an initiator DNA strand**.
- The enzyme required **Mg^{2+} ions**, as DNA synthesis was abolished in its absence.

3.3 Reverse Reaction: Pyrophosphorolysis Assay

- To assess whether the enzyme could **remove nucleotides**, it was incubated with **radiolabeled pyrophosphate (P^{32} -PPi)**.
- The enzyme **catalyzed nucleotide removal**, supporting the idea that it could **act dynamically in DNA metabolism**.

4. Significance of the Discovery

4.1 Identification of a New Class of DNA Polymerases

- Unlike **template-dependent polymerases**, this enzyme was **template-independent**, making it unique in DNA metabolism.
- It was later renamed **Terminal Deoxynucleotidyl Transferase (TdT)**.

4.2 Role in Immunology

- TdT is now known to play a **key role in immune system development**, particularly in **V(D)J recombination**.
- It **randomly adds nucleotides to DNA ends**, contributing to **antibody diversity** in B and T lymphocytes.

4.3 Applications in Molecular Biology

- TdT is now used for:
 - **End-labeling of DNA molecules** in research.
 - **Tail addition in genetic engineering**.
 - **TUNEL assay** (for detecting apoptotic DNA fragmentation).

5. Conclusion

- This study provided the **first evidence** for a **non-template-dependent** DNA polymerase, distinct from **Kornberg's DNA polymerase I**.
- The enzyme, now known as **Terminal Deoxynucleotidyl Transferase (TdT)**, catalyzes **random nucleotide addition** to the 3'-end of DNA.
- This work **expanded the understanding of DNA synthesis and repair mechanisms**, influencing fields like **molecular biology, immunology, and biotechnology**.